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Submission date: 15-Nov-2021 06:59PM (UTC+0700)

Submission ID: 1703343998

File name: 16_2019_flavonoid_on_ethanol_extract_rhizophora.pdf (542.97K)

Word count: 2794

Character count: 15433

ANALYSIS OF FLAVONOIDS ON ETHANOL EXTRACT OF RHIZOPHORA MUCRONATA FRUIT

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(Received 25 March, 2019; accepted 18 May, 2019)

Key words: Flavonoids, FT-IR, Mangrove, TLC-densitometry, Rhizophora mucronata

Abstract - Mangrove species $Rhizophora\ mucronata$ fruit has the potential to be utilized in a variety of processed foods. This study aimed to analyze the flavonoid compounds from the ethanol extract of fruit powder mangrove $Rhizophora\ mucronata$. The study conducted through several stages, extraction using ethanol solvent, phytochemical screening, FT-IR (Fourier Transform Infrared Spectroscopy), the creation of a standard solution and eluent preparation, isolation of flavonoids using TLC (Thin Layer Chromatography) densitometric method. The results of the analysis of phytochemical screening showed positive results containing flavonoids. Determination of Rutin at 0.32 to 0.46 Rf range obtained rutin an average grade $13.29\% \pm 0.43$. Infrared spectrum analysis showed absorption OH phenol in number $3260\ cm^{-1}$, C=C aromatic ring in the area from $1650\ to\ 1450\ cm^{-1}$, and the absorption at $1660\ cm^{-1}$ indicating a carbonyl group C=O.

INTRODUCTION

The world of mangrove ecosystems was reported in 124 countries located between 30^(North) and 30^(North) (South) latitude (Giesen et al., 2007; FAO, 2010). Closure of mangrove world estimated at 15,000,000 ha (FAO, 2010; Giri et al., 2011). The extent of mangrove forests in Asia (42%), followed by Africa (20%), North and Central America (15%), Oceania (12%) and South America (11%), while in Indonesia breadth is 3,112,989 ha which is a mangrove forest the widest in the world (Arobaya and Wanna, 2006; Giri et al., 2011). Mangrove forests are one of the tropical forests that store millions of benefits both ecologically, socio-economically, and social-cultural which are very important; for example, safeguarding coastal stability from abrasion, sources of fish, shrimp and other biodiversity, sources of fuelwood and building wood, and having functions of conservation, education, ecotourism and cultural identity (Setyawan, 2006). Mangrove species are often found in Asia, including in Indonesia, among others: Acanthus ebracteatus, Acanthus ilifolius, Acrostichum aureum, Bruguiera sp, Bruguiera hainessii and Rhizophora (Mesta et al., 2014; Umroh et al.,

2016).

One of the mangrove species found in Indonesia is Rhizophora mucronata a mangrove plants from the family Rhizophoraceae. These plants contain bioactive compounds such as alkaloids, carbohydrates, glycosides, tannins, proteins, and amino acids, flavonoids, saponins, sterols, acidic compounds, resins, peroxide, and polyuronides. Bioactive compounds are components derived from natural materials. Bioactive compounds used in the health sector among others to prevent cancer (Das et al., 2013), as antioxidants (Ravikumar, 2012), antibacterial (Ramasubburayan, 2015), antidiabetic (Hardoko, 2016), antimicrobial (Saad, et al., 2012) as well as anti-inflammatory (Chakraborty, 2016). One of the chemical constituents R. mucronata that is important is flavonoids. Flavonoids including one secondary metabolite (Harborne, 1984; Pietta et al., 2003; Kevin and Carolyne, 2006; Joel, 2010; Cahyani, 2016; Obidallah, 2018), which is uniformly dispersed in the vegetable kingdom, and includes one of the largest natural phenols.

Interest in food and pharmaceutical industries increased to flavonoids, as it has antioxidant compounds. Based on the literature search of *R*.

946 Ernawati*et al*

mucronata flavonoid compounds have been studied from the leaves (Behbahani et al., 2018), and roots (Asha, 2011), but for fruit, Rhizophora has not widely researched. The initial approach to determine the levels of flavonoids in fruits Rhizophora conducted qualitative phytochemical screening, followed by TLC detection and quantitative measurement of flavonoids using standard solutions quercetin and Rutin.

Rutin (quercetin-3-O-rutinoside) is a flavonoid commonly found in nature. At first, it was known as a mixture of flavonoids extracted from plants with the name "vitamin P" (Kalinova and Dadakova, 2009). Rutin can reduce capillary fragility and permeability, retinal hemorrhage and venous insufficiency (such as varicose veins, hemorrhoids, diabetic vascular disease) and improve vascular blood flow (tired legs, cramps) (Fathiazad et al., 2006). Rutin is also beneficial for the prevention and treatment of colorectal cancer and can prevent the pain induced by antitumor drugs (Azevedo et al., 2013). Currently, Rutin used as a single compound with chemical formulas, and its biological activity is essential, so it has the potential to be studied further. This study aims to analyze the flavonoid compounds of the ethanol extract of mangrove flour from R. mucronata using quercetin and rutin standards using the Thin Layer Chromatography (TLC)-densitomeric method.

MATERIALS AND METHODS

Collection and identification of fruits. Sample pieces of mangrove R. mucronata taken in the village of Kalianyar, Bangil, Pasuruan, East Java, Indonesia. Chemicals used was 96% ethanol, HCl, Magnesium, H₂SO₄, and other chemical materials that are reagent grade.

Preparation of fruit. This experiment started with the process of making flour *R. mucronata* includes sorting fruit, fruit leather stripping, immersion in fresh water and change water regularly until quite clear, boiling for 15 minutes, lower temperature drying between 50-70 °C to achieve 10-11% moisture, smoothing milled flour with tools Willey mill mesh sieve size 60. After that dried fruit flour for 10 minutes at a temperature of 70 °C, to prevent the powder into a dry and sour

Sample extraction. Extraction is done using solvent ethanol, *R. mucronata* fruit flour weighed 10 grams and supplemented with 96% ethanol, 250 mL.

Sample and the solvent extracted with speed extractor utilizing a pressure of 100 bar and a temperature of 70 °C, and 3x more repetitions to obtain the filtrate. Concentrated using a vacuum rotary evaporator temperature of 40 °C, 60 rpm, and a pressure of 200 mBar until no solvent drips.

Phytochemical screening. The preliminary phytochemical analysis of the plant extracts was performed using a standard protocol given by Harborne (1984) to identify the presence of flavonoids. Test flavonoid phytochemical active compound content of ethanol extract of *Rhizophora* done three times repetition. Extract *R. mucronata* put in a test tube is then dissolved in 1-2 mL of hot 50% methanol. After that, added Mg metal and 4-5 drops of concentrated HCl. The red or orange colored solution formed indicate flavonoids.

FT-IR analysis. Extract of R.mucronata then analyzed by FT-IR to measure the molecules of complex compounds.

Preparation of standard solution and eluent preparation. Raw carefully weighed 2mg quercetin (Sigma Q4951), then diluted with 10 mL of ethanol as a stock solution. Created dilution Quercetin with concentration of 0.100; 0.125; 0.150; 0.175 and 0.200 μg/ μL as the reference solution. The stationary phase Silica gel 60 F254 (Merck) with the sample volume of 1.0 μL and standard 1 μL. Eluent preparation for the determination of guercetin is Ethyl acetate: formic acid: Aquadest (85:10:15), while for the determination Rutin was Chloroform: Ethyl Acetate: Format acid (5: 4: 1). Distance elution 9 cm. After completion of the elution plate TLC is dried. Visual detection of the UV lamp 366 nm and detection wavelength TLC densitometry at 365 nm (Camag TLC Scanner 3). Interpretation of the sample contains Quercetin at Rf range from 0.85 to

Total flavonoid content measurement. Flavonoids total ethanol extract of *R. mucronata* sample calculated by linear regression of the calibration

0.90, and Rutin at Rf: 0.30 -0.40.

curve that has been measured quercetin previously. ${\tt RESULTS}$

Phytochemical screening and Determination of Quercetin levels using TLC-Densitometry

The test results showed that the ethanol extract of *R. mucronata* showed a positive reaction to the presence of flavonoids characterized by color changes to

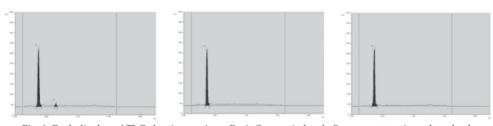


Fig. 1. Peak display of TLC-densitometric on Fruit Quercetin levels R. mucronata using ethanol solvent

orange. Results of analysis using standard flavonoid quercetin showed negative results in Figure 1.

The results of the ethanol extract of \overline{R} . mucronata Rf shown in Figure 2

The Rutin measurement results are shown in Table 1

Table 1. Rutin absorbance value at a wavelength of 366

Levels (μg/μL)	Spot Size (AUC)	
0.100	1,688.30	
0.125	2,745.20	
0.150	3,929.40	
0.175	4,787.80	
0.200	5,124.70	

Results of analysis using standard flavonoid quercetin showed negative results, then the results of the ethanol extract of R. mucronata using standard flavonoid quercetin showed Rf 0.34, 0.33 and 0.35. Based on the Rutin absorbance value in Table 1 has an equation: y = bx + a, where b = 35,661.60, a = 1,694.16, R2 = 0968, which is shown in Figure 3. After that equation obtained is used to determine the levels of Rutin found in mangrove fruit,

Analysis of FT-IR (Fourier Transform Infrared Spectroscopy)

The results of FT-IR spectrum shown in Figure 4

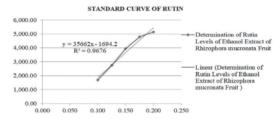


Fig. 3. Determination of standard curve Levels Rutin R. *mucronata* with ethanol

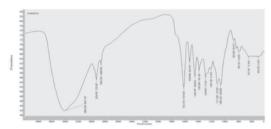


Fig. 4. FT-IR spectrum

DISCUSSION

Phytochemical screening and Determination of Quercetin and Rutin levels using TLC-Densitometry

Phytochemical screening analyzes performed on ethanol, which is polar. The test results showed that the ethanol extract of *R. mucronata* showed a positive reaction to the presence of flavonoids characterized

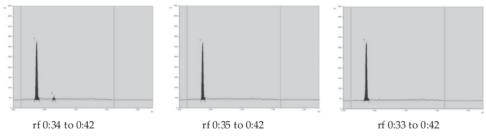


Fig. 2. Peak display of TLC-densitometric on Fruit Rutin levels R. mucronata using ethanol solvent

948 Ernawati*et al*

Table 2. Calculation Rutin levels R. mucronata fruit extracts in ethanol

Weighing (g)	Volume added (μL)	Area (AUC)	Content (%)	The mean (%)	standard deviation
10.00	3.00	3179.50	13.67	13.29	0.43
10.00	3.00	2874.90	12.81		
10.00	3.00	3076.80	13.38		

by color changes to orange. Ernawati *et al.* (2019) observed that fruit *R. mucronata* extracted with methanol also showed a positive reaction to the test as flavonoids, so it concluded that the flavonoids in the fruit test *R. mucronata* could be done using both types of such solvents.

Rutin levels of *R. mucronata* fruit ethanol extract produced a value of $13.29\% \pm 0.43$. According to Markham (1988), the levels of flavonoids and other phenolic compounds in plants vary between each part, tissue, and age of plants, and are influenced by environmental factors. These factors are temperature, UV-vis light, nutrition, water availability, and levels of CO₃ in the atmosphere.

Analysis of FT-IR (Fourier Transform Infrared Spectroscopy)

Infrared spectrum analysis showed the bands in the local catchment area number 3260.1660, 1620, 1520, 1440, 1365, 1285, 1260, 1225, 1200, 1175, 1145, 1125, 1080, 1040, 1010, 940, 860, 835, 780 and 745. Uptake in wave numbers 3260 cm-1 is OH absorption phenols that have hydrogen bonding (Silverstein et al., 1991). Bond types are shown in the catchment area 1300-800 cm-1 (C-C, C-O, C-N), 1900-1500 cm⁻¹ 1 (CP%O, CP%N, NP%O), 2300-2000 cm-1 (Ca"C, Ca"N), and 3000-2200 (C-H, O-H, N-H). Aromatic ring indicated by peaks that appear at 1650-1450 cm⁻¹ ¹region, isolated compounds providing a peak around 1620 cm⁻¹ and 1520 cm⁻¹ which is a strain of C=C aromatic and supported by an absorption band at 860 cm⁻¹, 835 cm⁻¹, 940 cm⁻¹ and in the area of 1440 cm-1 there is a very strong and sharp band which is an aromatic ring strain.

Isolated compounds showed absorption at wave 1660 cm cm⁻¹ which indicated the absorption of carbonyl group C = O, supported by a peak in 1145 cm⁻¹. According to the literature, the characteristics of C = O for flavonoid compounds are 1700-1750 cm⁻¹ which is supported by the presence of peaks in the wavenumber region 1158 cm⁻¹. Uptake of carbonyl compounds is smaller isolated because of their conjugation bond. The carbonyl compounds here are esters which are reinforced by peaks in the

region of 1300-1000 cm⁻¹ (Soliman *et al.*, 2017; Bakkialakshmi, 2017)

CONCLUSION

R. mucronata fruit extract was identified to contain flavonoids in Rutin standard solutions, characterized in FT-IR testing which result strain C = O wavelength 1660 cm⁻¹

ACKNOWLEDGEMENTS

We are grateful for the Minister of Research, Technology and Higher Education of the Republic of Indonesia, which has provided scholarships for studies Doctoral program.

REFERENCES

Arobaya, A. and Wanma, A. 2006. Tracing the rest of the mangrove forest area in Manokwari. Wetlands Conservation News. 14 (4): 4-5. [in Indonesian]

Asha, K.K., Suseela, M. and Lakshmanan, 2011. The flavonoids and phenolic compounds in two mangrove species and their antioxidant properties. *Indian Journal of Geo-Marine Sciences*. 41 (3): 259-264. Azevedo, M.I., Pereira, A.F., Nogueira, R.B., Rolim, F.E., Brito, G.A.C., Wong, D.V.T., five-Junior, R.C.P.,

Brito, G.A.C., Wong, D.V.T., five-Junior, R.C.P., Ribeiro, R.A. and Vale, M.L. 2013. The antioxidant effects of the flavonoids quercetin inhibit Rutin and chronic oxaliplatin-induced painful peripheral neuropathy. *J Molecular Pain*. 9 (53): 1-14.

Bakkialakshmi and Jayoti, R. 2017. Some infrared spectrum analysis of flavonoids with hemoglobin. Int.J of Applied and Advanced Scientific Research (IJAASR). 2 (2): 2456-3080.

Behbahani, B.A., Farideh, T.Y., Fakhri, S., Hamid, N., Alireza, V. and Ali, A. 2018. Phytochemical analysis and antibacterial activities of mangrove leaf extracts against the growth of some pathogenic bacteria. *J Microbial Pathogenesis*. 114: 225-232.

Cahyani, Maulidya, D. and Novidayasa, 2016. Natural Dyes Extraction of Mangrove Wood (*Rhizophora mucronata*) Microwave Assisted Extraction Method. [Thesis]. Institute of Technology, Surabaya [Indonesian]

Chakraborty, K. and Vamshi, R. 2016. Two rare antioxidant and anti-inflammatory oleananes from loop root

- Asiatic mangrove Rhizophora mucronata. Phytochemistry. XXX: 1-9
- Das, S. and Abhay, K.P. 2013. Chemistry and biological activities of flavonoids: an overview. *The Scientific World Journal*. 1-16.
- Ernawati, E. Suprayitno, Hardoko, and Yanuhar, U. 2019.

 Extraction of bioactive compounds from *Rhizophora mucronata* fruit using sonication method. *Proceeding of The 1st International Conference on Fisheries and Marine Science*. Airlangga University, Surabaya. *IOP Conf. Series: Earth and Environmental Science*. 236: 012122.
- FAO Forestry, 2010. Global Forest Resources Assessment. Food and Agriculture Organization of the United Nations. Rome, Italy.
- Fathiazad, F., Delazar, A., Amiri, R. and Sarker, S.D. 2006. Extraction of flavonoids and quantification of rutin from a waste of tobacco leaves. *Iranian J. Pharm. Res.* 3: 222-227.
- Giesen, W., Wulffraat, S., Zieren, M. and Scholten, L. 2007.
 Mangrove Guidebook for Southeast Asia. FAO
 Regional Officer for Asia and the Pacific. Bangkok,
 Thailand.
- Giri, C., Ochieng, Tieszen, L.L., Zhu, Z., Singh, A., Loveland, T., Masek, J. and Duke, N. 2011. Status and distribution of mangrove forests of the world using earth observa.tion satellite data. *J Global Ecology* and Biogeography. 20: 154-159.
- Harborne, 1984. Phytochemicals Methods, A Guide to Modern Techniques of Plant Analysis. 2nd edition. Chapman and Hall. USA.
- Hardoko, Eddy S., Puspitasari and Amalia, 2016. Study of *Rhizophora mucronata* ripe fruit flour as a functional food for antidiabetic. *International Food Research Journal*. 22 (3): 953-959.
- Joel, E.L. and Valentin, B. 2010. Isolation and characterization of secondary metabolites from the mangrove plant Rhizophora mucronata. Asian Pacific Journal of Tropical Medicine. 602-604.
- Kalinova, J. and Dadakova, E. 2009. Rutin and total quercetin content in Amaranth (*Amaranths* spp.). *Plant Foods Hum. Nutr.* 64: 68-74.

- Kevin, S., Gould and Carolyn, L. 2006. Flavonoids functions in plants. In: Andersen, Kenneth M (eds) Flavonoids: Chemistry, Biochemistry, and Applications. Taylor & Francis CRC
- Markham, K.R. 1988. Techniques of Flavonoid Identification. London: Academic Pr.
- Mesta, P.N., Setturu, B., Subash, C.M.D., Rajan, K.S. and Ramachandra, T.V.2014. Inventorying, mapping and monitoring of mangroves towards sustainable management of West Coast, India. J Geophys Remote Sensing. 3 (3).
- Pietta, P., Claudio, G. and Annamaria, P.2003. Flavonoids in herbs. In: C Evans (ed) *Flavonoids in Health And Disease*. Marcel Dekker Inc. 2nd edition.
- Obidallah, Aljaghthmi, Hassaan, H. and Isam, A.Z. 2018. Bioactive compounds extracted from mangrove plants (*Avicennia marina* and *Rhizophora mucronata*): an overview. *Pathophysiology*. 14: In Press.
- Ravikumar, S. and Gnanadesigan, M. 2012. Hepatoprotective and antioxidant properties of mangrove plant Rhizophora mucronata in CCl₄ Intoxicated Rats. Journal of Experimental & Clinical Medicine. 4 (1): 66-72.
- Saad, S., Muhammad, T., Deny, S., Haitham, Q. and Anis, F.I. 2012. *In vitro* antimicrobial activity of mangrove plant Sonneratia alba. *Asian Pacific Journal of Tropical Biomedicine*, 427-429.
- Setyawan, A.D. and Kusumo, W. 2006. Conservation problems of mangrove ecosystem in coastal area of Rembang Regency, Central Java. *Biodiversitas*. 7 (2): 159-163.
- Silverstein, R.M., Clayton, B.G. and Torence, C.M. 1991. Spectrometric Identification of Organic Compounds. John Wiley & Sons, Inc.
- Soliman, Ahmed, M.B., Ahmed, H.M. and Mohamed, A.F.H. 2017. Antifouling evaluation of extracts from Red Sea soft corals against primary biofilms and biofouling. Asian Pacific Journal of Tropical Biomedicine. 1-7.
- Umroh, Wahyu, A. and Holy, P.S. 2016. Detection of mangrove distribution in Pongok Island. *Procedia Environmental Sciences*. 33: 253-257.

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PAGE 1	
PAGE 2	
PAGE 3	
PAGE 4	
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